

Organocobaloximes: synthesis, oxygen insertion and kinetics

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Abstract

A total of 27 organocobaloximes have been synthesised and characterized with three different dioxime ligands, dmgH, chgH and dpqH. Many of the chgH and all the dpqH complexes have been synthesised for the first time. The insertion of molecular oxygen into these organocobaloximes, $\text{RCo}^{\text{III}}(\text{L}_2)\text{B}$ [$\text{L} = \text{dmgH}, \text{chgH}$ and dpqH] under thermal and photochemical conditions have been studied. Kinetic studies at ambient temperature under irradiation show that the rate of insertion depends upon the nature of R, L, B and the solvent. The rate follows the order $\text{dpqH} > \text{chgH} > \text{dmgH}$; Naphthyl > heteroaromaticmethyl > benzyl; piperidine > morpholine > γ -picoline > pyridine > 2-bromopyridine > 2-acetylpyridine; and the rates are faster in acetonitrile than in acetone and chloroform. In methanol, the benzylic cobaloximes exist as an equilibrium mixture of solvated penta-coordinated and hexa-coordinated species and in chloroform these exist as hexa-coordinated species. © 1998 Elsevier Science S.A. All rights reserved.

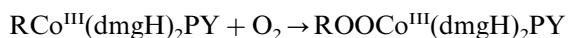
Keywords: Organocobaloximes; Dioxygen insertion reactions; Kinetic studies

1. Introduction

Organocobaloximes, $\text{RCo}^{\text{III}}(\text{dmgH})_2\text{B}$ (where R is an organic group, B is the coordinating base) have been extensively studied [1], ever since Schrauzer first highlighted their importance as models of coenzyme B_{12} [2]. Since then it has outgrown its initial relevance and has acquired an independent research field because of its rich chemistry [3]. The key feature of these complexes is the weak Co–C bond ([4]a,b) and this bond in alkyl cobaloximes has been reported to be homolytically cleaved by irradiation with visible light in a manner similar to the activation of vitamin B_{12} by apoenzyme ([4]c,d,e). These compounds find a great use in organic synthesis [5] and in catalysis [6]. Since small structural changes in the molecule have a profound effect on its reactivity, the synthesis of new organocobaloximes continues to attract the attention of the chemists [1,3,7]. The importance of organocobaloximes with ligands

other than dmgH has recently been perceived in many reactions in organic synthesis [7,8].

We are, in particular, interested in the reaction of organocobaloximes with molecular oxygen. Reactions involving fixation of O_2 are important not only for a biological system but also for some synthetic processes ([9]a–e). Some reactions of alkylcobaloximes have been reported, e.g. O_2 has been shown to insert into Co–C bond to give a stable 1:1 dioxy adduct ([9]f,g,h).



Both thermal and photochemical activation of these reactions have been observed. The reaction conditions vary over a wide range depending on the nature of R group, for example, when R is benzyl or allyl the reaction proceeds both under thermal as well as photochemical conditions but when R is alkyl, the photochemical conditions are essential [10]. Stereochemical studies have confirmed that such reactions occur with racemisation at the α carbon [11]. The last review on the insertion of oxygen in to M–C bond giving peroxo complexes was published by Wojcicki [12].

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Table 1

The yield, spectroscopic and R_f data for $RCH_2Co^{III}(L_2)Py$ and $RCH_2OOCo^{III}(L_2)Py^a$

Composition	Yield (%) ^b	¹ H-NMR chemical shifts, δ (ppm), CDCl ₃						λ_{max} (nm) ^c	R_f^d	
		Aromatic	CH ₂ Co (s)	L	Pyridine					OH...H
					α	γ	β			
2a		6.87–7.21	4.31	2.21,2.31	8.38	7.62	7.28	17.81	0.18	
3a		6.87	4.31	2.21, 2.31	8.34	7.62	7.28	17.81	0.17	
4a		6.65–7.20	4.31	2.15, 2.25	8.35	7.59	*	18.66	0.17	
5a		6.73–7.36	4.31	2.28, 2.38	8.41	7.54	*	18.00	0.16	
9a		7.00–7.97	4.86	2.21	8.48	8.08	*	17.81	0.11	
10	74	6.67–7.12	2.90	1.46 2.48,	8.60	7.62	7.38	17.81	245, 466.7	0.62
10a		6.88–7.50	4.35	1.57, 2.80	8.39	7.65	*	—	0.18	
11	71	6.59–7.20	2.80	1.51, 2.50	8.52	7.69	7.25	17.69	244, 464.8	0.59
11a		6.88–7.38	4.31	1.60, 2.82	8.38	7.71	*	0.18		
12	76	6.88–7.50	2.76	1.52, 2.51	8.62	7.66	*	17.69	231, 461.6	0.58
12a		6.88–8.12	4.37	1.59, 2 71	8.35	*	*	0.18		
13	62	6.60–6.99	2.91	1.47, 2.49	8.53	7.53	7.28	240, 468.0	0.59	
13a		6.56–7.50	4.38	1.56, 2.88	8.12	7.81	*	0.18		
14	64	6.25–7.06	2.87	1.47, 2.49	8.56	7.71	7.29	17.19	243.5, 466.7	0.63
14a		6.56–7.52	4.38	2.88	8.12	7.81	*	0.20		
18	72	6.88–8.12	3.40	1.26, 2.29	8.57	*	*	17.94	231, 467.2	0.60
18a		6.80–8.75	5.00	1.62, 2.81	9.06	*	*	0.18		
19	70	6.71–7.68	3.55	*	8.97	7.87	*	18.68	244, 472.9	0.34
19a		6.84–7.55	4.71	*	8.84	7.87	*	18.90	0.14	
20	75	6.64–7.48	3.36	*	8.87	7.74	*	18.68	236.5, 471.9	0.34
20a		7.19–7.27	4.55	*	8.72	7.42	*	18.75	0.14	
21	72	6.58–7.55	3.35	*	8.77	7.81	*	18.68	229, 472.5	0.30
21a		7.16–7.24	4.57	*	8.67	7.76	*	18.72	0.10	
22	71	7.00–7.81	3.41	*	8.87	7.81	*	18.74	261.5, 472.6	0.30
22a		7.16–7.81	4.63	*	8.67	7.74	*	18.76	0.11	
23	70	6.76–7.50	3.41	*	8.81	7.72	*	18.74	245, 472.6	0.30
23a		7.60–7.81	4.64	*	8.67	7.61	*	18.76	0.10	
24	78	6.71–7.61	3.68	*	8.97	7.81	*	18.62	245	0.36
24a		6.77–7.50	4.84	*	8.77	7.80	*	18.88	0.16	
25	76	6.64–7.68	3.55	*	8.97	8.01	*	18.75	242.5, 472.4	0.36
25a		7.17–7.25	4.60	*	8.69	7.75	*	18.68	0.16	
26	78	6.77–7.74	3.32	*	8.85	7.87	*	18.75	244.5	0.31
26a		7.21–7.42	4.54	*	8.68	7.75	*	18.75	0.11	
27	70	6.66–8.00	4.03	*	8.92	8.02	*	18.38	226sh, 472.5	0.30
27a		6.84–7.81	5.03	*	8.74	7.87	*	18.65	0.10	

^a All the complexes give the satisfactory elemental analysis.^b Yields for the inserted products are almost quantitative in each case. The known compounds are not listed, ([9]f, [30]).^c The first value refers to MLCT and the second one to Co–C CT band.^d Ethylacetate is the eluent for the compounds **1–18** and **1a–18a** and a mixture of EtOAc:CCl₄ (1:9) is used for the compounds **19–27** and **19a–27a**.^e Merge with aromatic.

gen. (E) The rate of reaction is slowed down in the presence of radical inhibitor like galvinoxyl; (F) the rate of insertion changes as the equatorial ligand is changed and it follows the order $dpgH > chgH > dmGH$ in all cases. (G) The solvent affects the rate and it follows the order acetonitrile > acetone > chloroform; (H) the reaction proceeds faster as the axial base strength increases; 2-acetylpyridine < 2-bromopyridine < pyridine < γ -picoline < morpholine < piperidine; (I) the rate changes with the change in R group and it follows the order naphthyl > heteroaromaticmethyl > benzyl for the three series of complexes. Among the

benzyls, *p*-methoxybenzyl has the highest rate and among the heteroaromaticmethyl groups it follows the order 2-thienylmethyl > 3-thienylmethyl > furfuryl for all the three series of complexes. It seems clear that the rate of insertion depends upon various factors including the nature of R, L, B and the solvent.

Oxygen insertion into M–C bond is a reaction which occurs with remarkably a wide range of organometallic compounds. Both thermal as well as photochemical activation of these insertions have been observed in the Co–C bond of organocobalt complexes. The results in the photochemical reactions are much more difficult to

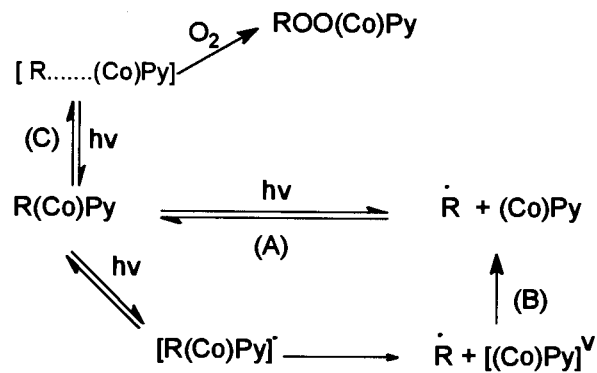
interpret owing to the greater complexity of this reaction compared to thermal reactions; the exact role of light in these reactions is not known. Many suggestions have been made by various workers, e.g. Schrauzer has proposed, based in part on MO calculations, that photolysis of alkyl cobaloximes leads to excitation and subsequent homolytic cleavage of the Co–C bond [17]. Giannotti and coworkers have suggested that photolysis with light in the charge transfer region (about 450 nm) leads to the expulsion of the axial base ligand as the primary photochemical process [18]. The Co–B bond is reformed after the insertion has taken place.

Several mechanisms including a radical chain mechanism can be envisaged for the insertion of oxygen into Co–C bond in organocobaloximes. Such radical chain reactions generally require induction period and once these start they proceed to completion without the external stimulation by light. However, in view of the experimental observations in the present study that (a) there is no induction period; (b) the reaction stops as soon as the light stimulation is stopped; (c) the organocobaloximes in the absence of oxygen are stable at 30°C, i.e. at a temperature where the insertion occurs readily as shown by kinetics, we rule out the participation of such a chain mechanism.

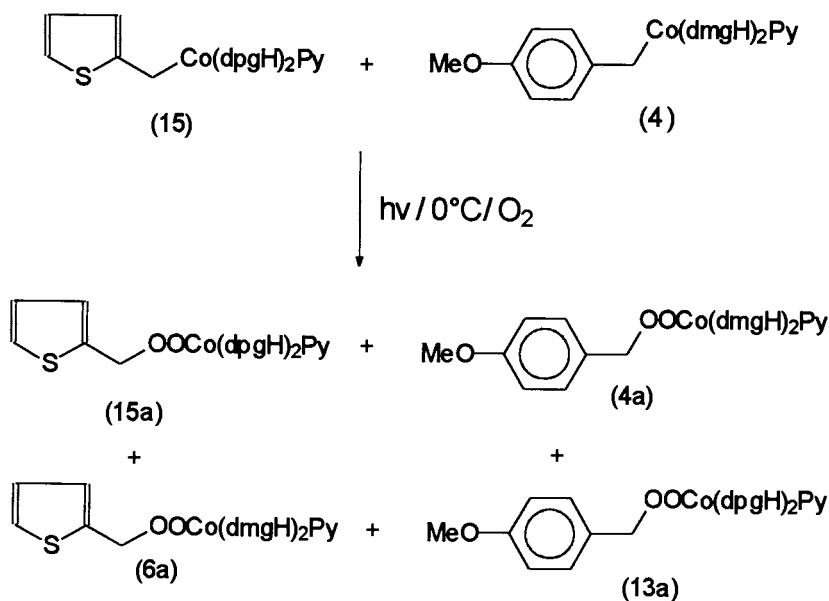
We have not considered any mechanism involving thermal conditions since no insertion has been observed in the dark under thermal conditions. Similarly, any mechanism which involves the loss of pyridine at any stage in the photochemical process has also been ignored. Even if such a base off process was operating then the rate of insertion should have increased with

the decrease in base strength, however a reverse order has been observed in the present study.

Among the three other possibilities A, B and C, the basic difference in these mechanisms is that in (A), the free radicals R and (Co)Py are involved. R picks up O₂ and the adduct combines with (Co)Py to give the insertion product. In this the stability of R plays an important role and should affect the rate. Mechanism (C) on the other hand does not involve a complete rupture of Co–C bond, the resulting radical pair (R...Co) within the solvent cage on reaction with O₂ gives the final inserted product. In this case the nature of solvent should affect the rate. Such a mechanism has been proposed by many workers [19–21].



It seems difficult to distinguish between these possibilities without doing a more detailed studies but since the solvent and the R group do affect the rates in these reactions, it looks certain that the mechanism (A) and (C) are operative. It is, however, very difficult to know their relative contribution to the overall rate. The formation of cross insertion products further suggest that a complete rupture of bond takes place, i.e. the mechanism (A) is operating.



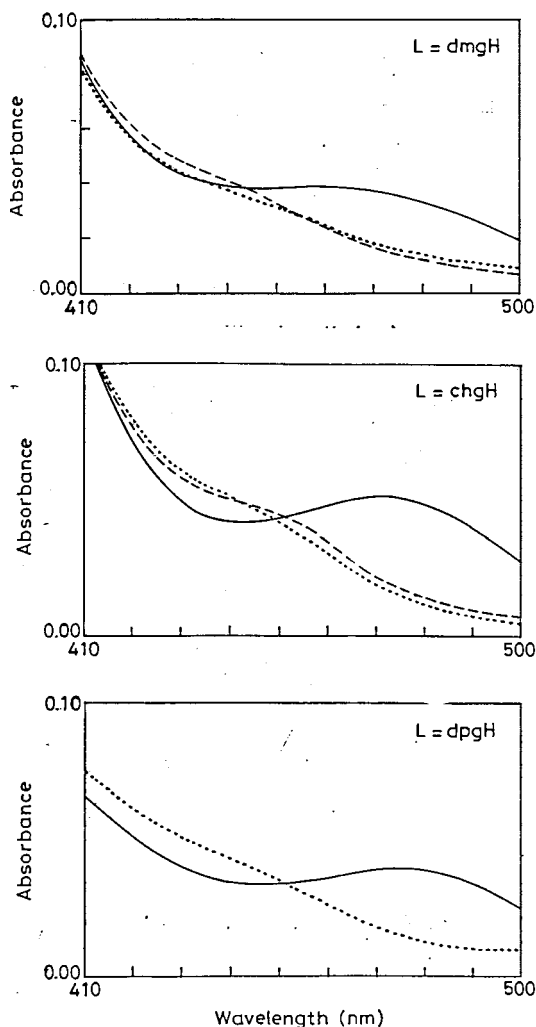


Fig. 1. Effect of the addition of pyridine (— 0.0 M, ---- 10^{-3} M, ... 10^{-2} M saturation curve) to a 10^{-5} M solution of benzyl(L)₂Py in methanol (L = dmgH (1), chgH (10); for the complex L = dpqH (19), the plot is only qualitative.

Finally, the change in the insertion rate for a given compound having the same R and B but different equatorial ligand L is hard to explain. One of the contributing factors may be the *cis* influence exerted by the bulkier equatorial ligand on the Co–C bond. We find that the magnitude of Δi_p is significantly larger [0.56–0.92] ppm than Δi_c [0.00–0.34] ppm in all cases where $\Delta i_p = \delta i_{dpqH} - \delta i_{dmgH}$ and $\Delta i_c = \delta i_{chgH} - \delta i_{dmgH}$ and δi is the chemical shift. If this is taken to be as the extent of *cis* influence, as described in earlier papers ([7]b, [16,22]), then the order of *cis* influence follows the order $dpqH > chgH \cong dmgH$. The extent of *cis* influence is felt most on the Co–CH₂ followed by P_z.

3. Experimental

Chlorocobaloxime, ClCo^{III}(L₂)Py (L = dmgH, chgH and dpqH) [15]. 2-Chloromethylthiophene, 3-bromomethylthiophene, furfurylbromide, 4-cyanobenzylbromide, 4-methoxybenzylbromide, 3-methoxybenzylbromide were synthesised according to the published procedures [23–28]. Diphenylglyoxime, 1,2-cyclohexanedionedioxime and all the other halides were obtained from Aldrich and were used as such.

The organocobaloximes were synthesised by a general procedure detailed earlier in many papers ([7]b, [15]) and involves the reaction of cobaloxime(I), with the corresponding benzylic halide. Cobaloxime(I) was prepared in situ by the NaBH₄ reduction of chlorocobaloximes in aqueous methanol under strict oxygen free atmosphere. The work-up procedure was similar to the ones described earlier ([7]b, [15]). The crude products were purified by column chromatography as described earlier for alkyl cobaloximes [15].

2-ThienylmethylCo^{III}(dpqH)₂B [B = 2-acetylpyridine, 2-bromopyridine, picoline, morpholine, piperidine] were prepared by replacing the aqua group from 2-thienylmethylCo^{III}(dpqH)₂H₂O. In a typical experiment morpholine (0.029, 0.23 mmol) in acetone (1 ml) was added dropwise to a solution of 2-thienylmethyl(aquo)cobaloxime (0.129, 0.18 mmol) in acetone (3 ml). The reaction mixture was stirred in dark in an inert atmosphere. The completion of the reaction was checked with TLC on silica gel using ethyl acetate:CCL₄ in 1:9 ratio. The wine red coloured solution fumed orange within minutes and the orange precipitate which formed were filtered, washed with acetone (2 × 2 ml) and dried in vacuum over P₂O₅. Yield 68%

The aqua complex was prepared from the corresponding pyridine complex following the literature procedure [29]. In a typical experiment, 2-thienylmethylCo^{III}(dpqH)₂Py (214 mg 3 mmol) was dissolved in acetone (30 ml). To this was added 5 ml of water and 248 mg of Dowex 50w-x ion exchange resin (H⁺ form) and the mixture was stirred for 16h in the dark. The mixture was filtered and the filtrate was evaporated to 510 ml, cooled in ice and filtered to produce orange powder which was washed with cold water and dried in vacuo over P₂O₅. Yield 80%

3.1. Physical measurements and instruments

The ¹H-NMR spectra were recorded on a Bruker WP-80 FT NMR machine. UV-vis absorption spectra were recorded on Shimadzu UV-190 spectrometer at ambient temperature. The elemental analysis were undertaken at the Regional Sophisticated Instruments Centre, Lucknow.

3.2. General procedure for oxygen insertion under photochemical conditions

In a typical experiment a solution of organocobaloxime (0.2 mmol in 100 ml dichloromethane) at 0°C was irradiated with 2 × 200 W tungsten lamps kept at a distance of ca. 10 cm. and pure oxygen was bubbled into this solution. The progress of the reaction was monitored by TLC on silica gel using ethylacetate (dmgH and chgH) or 1:9 ethylacetate/CCl₄ mixture (dpgH) as the eluent. The reaction was complete within 2 h in all cases. There was a distinct colour change from yellow-orange to dark brown at this stage. At the end of reaction, the solvent was stripped off and the crude product was purified on silica gel column using ethyl acetate as the eluent. The yields were almost quantitative in all cases.

3.3. Oxygen insertion in presence of 2-acetylpyridine

The same general procedure as above was followed except that 2-acetylpyridine (3–4-fold excess) was added to the reaction mixture. After the completion of the reaction, the solvent was completely evaporated and the solid was washed three to four times with diethylether. The ¹H-NMR on the residue showed that it had no 2-acetylpyridine. The ether on evaporation gave back all the 2-acetylpyridine.

3.4. Cross insertion reaction between 2-thienylmethyl dpgH complex (15) and 4-methoxybenzyl dmG complex (4) in dichloromethane at 0°C

The general procedure is same as outlined above. In a typical experiment, a solution of two organocobaloximes 2-thienylmethyl dpgH complex (15, 0.424g, 0.59 mmol) and 4-methoxybenzyl dmG complex (4, 0.291g, 0.59 mmol) in 300 ml dichloromethane was irradiated with 2 × 200 W tungsten lamps at 0°C. After completion of the reaction the semi-solid mixture, dissolved in minimum amount of chloroform, was loaded on silica gel column (100–200 mesh) pre-eluted with chloroform. The polarity of the solvent was carefully increased with ethylacetate (0–10%). The first fraction contained organic product followed by dpgH complexes and finally the dmG complexes.

3.5. Kinetics of oxygen insertion under photochemical conditions at 30°C procedure

Pure oxygen was bubbled through chloroform or methanol (100 ml) for 2 h and a solution of organocobaloxime (3–4 mg) was made (10–5 M) in the above solvent. This solution was taken in the spectrophotometric cell, stoppered and was irradiated with

200 W tungsten lamp at 30°C. The progress of the reaction was monitored by measuring the absorbance at 420 nm at intervals.

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